

VASO-OCCLUSIVE DEVICES WITH BIOACTIVE ELEMENTS

Field of Invention

[0001] The invention pertains to medical devices, and more particularly to vaso-occlusive devices with internal biologically active agents.

5 Background

[0002] In many clinical situations, blood vessels are occluded for a variety of purposes, such as to control bleeding, to prevent blood supply to tumors, and to block blood flow within an aneurysm, arteriovenous malformation, or arteriovenous fistula.

10 [0003] Embolization of blood vessels is particularly useful in treating aneurysms. Aneurysms are abnormal blood filled dilations of a blood vessel wall, which may rupture causing significant bleeding. For the cases of intracranial aneurysms, the significant bleeding may lead to damage to surrounding brain tissue or death. Intracranial aneurysms may be difficult to treat when they are
15 formed in remote cerebral blood vessels, which are very difficult to access. If left untreated, hemodynamic forces of normal pulsatile blood flow can rupture fragile tissue in the area of the aneurysm causing a stroke.

[0004] Vaso-occlusive devices have been used in the treatment of aneurysms. Vaso-occlusive devices are surgical implants placed within blood
20 vessels or vascular cavities, typically by using a catheter as a conduit, to arrest blood flow, form a thrombus and occlude the site. For instance, a stroke or other such vascular occurrence may be treated by placing a vaso-occlusive device

proximal of the site to block the flow of blood to the site and alleviate the leakage.

An aneurysm may similarly be treated by introducing one or more vaso-occlusive devices through the neck of the aneurysm. The placement of the vaso-occlusive device(s) helps cause a mass to form in the aneurismal sac and alleviate the

5 potential for growth of the aneurysm and its subsequent rupture. Other diseases, such as tumors, may often be treated by occluding the blood flow to the tumor.

[0005] There are a variety of known vaso-occlusive devices suitable for creating an embolic obstruction for therapeutic purposes. One such device is found in U.S. Patent No. 4,994,069, to Ritchart et al.. That patent describes a
10 vaso-occlusive coil that assumes a linear helical configuration when stretched and a folded convoluted configuration when relaxed. The coil has a stretched configuration when placed in a catheter, which is used in placement of the coil at the desired site, and assumes the convoluted configuration when the coil is ejected from the catheter and the coil relaxes. Ritchart et al. describes a variety
15 of shapes, including "flower" shapes and double vortices. A random shape is described as well.

[0006] Vaso-occlusive coils having complex, three-dimensional structures in a relaxed configuration are described in U.S. Patent No. 6,322,576B1 to Wallace et al. The coils may be deployed in the approximate shape of a sphere, an ovoid, a
20 clover, a box-like structure or other distorted spherical shape. The patent also describes methods of winding the anatomically shaped vaso-occlusive device into appropriately shaped forms and annealing them to form various devices.

[0007] Vaso-occlusive coils having little or no inherent secondary shape have also been described. For instance, U.S. Patent Nos. 5,690,666 and 5,826,587 both by Berenstein et al. describe coils having little or no shape after introduction into the vascular space.

5 [0008] It is known to coat vaso-occlusive devices with a bioactive material that enhances a thrombogenic characteristic of the device, or that promotes conversion of thrombus to cellular tissues. For example, U.S. Patent No. 6,280,457B1 to Wallace et al., describes an occlusive device including an inner core wire covered with a polymeric material. The polymeric material includes
10 protein based polymers, absorbable polymers, non-protein based polymers, and combinations thereof. The polymer facilitates the processes of thrombosis within a body cavity and/or conversion of thrombus into dense cellular tissue to stabilize the occlusion of a body cavity. However, the coating of bioactive material may increase friction between the occlusive device and an occlusive device delivery
15 tool during deployment of the occlusive device. In some cases, the coating may even cause the occlusive device to adhere to the delivery tool or to a packaging. The coating of bioactive material may also alter a mechanical behavior of the occlusive device.

[0009] One problem associated with existing vaso-occlusive devices is that
20 they may not have a sufficient strength or stiffness to retain their shape after they are delivered into an aneurysm. When the above-mentioned vaso-occlusive devices are placed within an aneurysm, they tend to induce a formation of a

thrombi for occlusion of the aneurysm. However, with time and the influence of hemodynamic forces and thrombolytic processes, the delivered vaso-occlusive devices may move or change shape due to their relatively low stiffness or through the process of seeking a minimally energetic morphology. As a result, 5 the delivered vaso-occlusive devices may move out of the position or shape in which they were originally placed. In some cases, the delivered vaso-occlusive devices may even dislodge out of the sack of an aneurysm.

SUMMARY OF THE INVENTION

[0010] In accordance with one aspect of the invention, a vaso-occlusive device having an agent delivery capability is provided. In one embodiment, the vaso-occlusive device includes a coil and an agent carrier disposed within a lumen of the coil. The agent carrier includes a bioactive material or agent that elicits a tissue reaction when placed inside a body. By way of non-limiting examples, the agent carrier can have an elongate shape, be in a form of a sphere, a cone, a plate, a mesh, or other customized shape.

[0011] The agent carrier can be made from a biodegradable material, in which case, the composition of the agent carrier includes a bioactive material or agent that is released when placed inside a body. The agent carrier can also be made from a non-biodegradable material, in which case, the bioactive material or agent is coated onto a surface of or incorporated within the agent carrier. In other embodiments, the agent carrier is made from a material that adheres or absorbs a bioactive agent. By way of non-limiting examples, the agent carrier can include one or more polymer filaments, a sponge, a tube, a cloth, or other materials that are capable of encompassing, absorbing or adhering a bioactive agent. In this case, the agent carrier is used to deliver the bioactive agent, which will diffuse out of the agent carrier into the surroundings when placed in a target site. One advantage of this embodiment is that placing the agent carrier within the lumen of the coil allows an exterior of the coil to be unaffected by the bioactive material during delivery of the coil. That is, the bioactive material would not increase a

friction between the coil and a delivery tool, and would not cause the coil to be adhered to the delivery tool or to a packaging.

[0012] Other aspects and features of the invention will be evident from reading the following detailed description of the preferred embodiments, which

5 are intended to illustrate, not limit, the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

[0013] The drawings illustrate the design and utility of preferred embodiments of the present invention, in which similar elements are referred to by common reference numerals, and in which:

5 [0014] **FIG. 1** is a side view of a vaso-occlusive device in accordance with one embodiment, including an agent carrier disposed within a lumen of a coil;

[0015] **FIG. 2** is a side view of a vaso-occlusive device in accordance with another embodiment, including an active element disposed within a lumen of a coil;

10 [0016] **FIG. 3** is a side view of the vaso-occlusive device of **FIG. 2**, showing the active element having an expanded configuration;

[0017] **FIGS. 4-6** are side views of embodiments of vaso-occlusive devices;

[0018] **FIGS. 7-13** show embodiments of vaso-occlusive devices having secondary shapes;

15 [0019] **FIG. 14** is a cross-sectional side view of an embodiment of a vaso-occlusive device being delivered using a delivery catheter, showing the coil of the vaso-occlusive device having a substantially rectilinear shape inside the delivery catheter;

[0020] **FIG. 15** is a cross-sectional side view of an embodiment of a vaso-
20 occlusive device being delivered using a delivery catheter, showing the vaso-occlusive device changing from a stretched configuration to relaxed configuration as it exits from the delivery catheter;

[0021] **FIG. 16** is a side view of a portion of a delivery catheter from which a vaso-occlusive device is deployed and mechanically released; and

[0022] **FIG. 17** is a side view of a portion of a delivery catheter from which a vaso-occlusive device is deployed and electrolytically released.

[0023] DETAILED DESCRIPTION OF ILLUSTRATED EMBODIMENTS

[0024] Delivery of a Bioactive Agent

[0025] In accordance with one aspect of the invention, the vaso-occlusive device 10 of **FIG. 1** is provided with an agent carrier 14 carried by the coil 12.

5 The coil 12 is made from a linear element 16, such as a wire, which preferably has a circular cross-sectional shape. In alternative embodiments, the linear element 16 of the coil 12 may have a rectangular, triangular, other geometric cross-section, or an irregular shaped cross-section. The coil 12 includes one or more loops or windings 18 formed by the linear element 16. The loops 18 define
10 a central lumen 20 in which the agent carrier 14 is placed. In the illustrated embodiment, the vaso-occlusive device 10 has an overall diameter or cross-section which is preferably in the range of 0.010 to 0.023 inches. However, the vaso-occlusive device 10 may have other diameters and/or cross-sections, as well.

15 [0026] The vaso-occlusive device 10 may optionally include one or more end caps 22 secured to a first end 24 or to a first and a second end 26 of the coil 12.

[0027] The coil 12 may have an open or closed (e.g., **FIGS. 9-11**) pitch. The coil 12 shown in **FIG. 1** may be constructed by wrapping the linear element 16, such as a wire, around a mandrel, stylet, or other shaping element. The coil 12
20 may optionally be heat treated, as known to one skilled in the art. It should be noted that the formation of vaso-occlusive devices having a helical coil shape is well known in the art, and need not be described in further detail.

[0028] The coil 12 may be made of a variety of materials, such as metals or polymers. Suitable metals and alloys for the coil 12 may include the Platinum Group metals, especially platinum, rhodium, palladium, rhenium, as well as tungsten, gold, tantalum, and alloys of these metals. These metals have significant radiopacity and their alloys may be tailored to accomplish an appropriate blend of flexibility and stiffness. These metals are also largely biologically inert. The coil 12 may also be formed from stainless steels if some sacrifice of radiopacity may be tolerated. Other materials that may be used may include "super-elastic alloys," such as nickel/titanium ("Nitinol") alloys, copper/zinc alloys, or nickel/aluminum alloys. Exemplary alloys that may be used are described in U.S. Patent Nos. 3,174,851, 3,351,463, and 3,753,700. If Nitinol is used, the diameter of the coil 12 may be significantly smaller than that of a coil 12 made from relatively more ductile platinum or platinum/tungsten alloy.

[0029] Examples of polymers that may be used for construction of the coil 12 includes polydienes, polyalkenes, polystyrenes, polyoxides, polycarbonates, polyesters, polyanhydrides, polyurethanes, polyamides, polyimides, polyacrylics, polymethacrylics, polyacetals, and vinyl polymers. The coil 12 can alternatively be made of radiolucent fibers or polymers, such as Dacron (polyester), polyglycolic acid, polylactic acid, fluoropolymers (polytetrafluoroethylene), Nylon (polyamide), and/or silk.

[0030]. If the coil 12 is not made from a radiopaque material, the coil 12 may be coated, mixed, or filled with radiopaque materials such as metals (e.g.

tantalum, gold, tungsten or platinum), barium sulfate, bismuth oxide, bismuth subcarbonate, zirconium oxide, and the like. Alternatively, continuous or discrete radiopaque markers may be incorporated within or affixed to the coil 12.

[0031] As shown in **FIG. 1**, the agent carrier 14 includes one or more axially oriented elements 30 having a substantially rectilinear or a curvilinear (less than 360°) configuration along a length of the vaso-occlusive device 10. In the case of a more complex coil shape, the active element could mirror the shape of the coil. The axially oriented element 30 is located within the lumen 20 of the coil 12 and is secured to the ends 24 and 26 or the end caps 22 of the coil 12. The securing may be accomplished by an anchor or a suitable adhesive, such as ultraviolet-curable adhesives, silicones, cyanoacrylates, or epoxies. Alternatively, the axially oriented element 30 can be secured to the coil 12 by chemical bonding between reactive groups on the axially oriented element 30 and the coil 12, solvent bonding, fusing both materials so that they melt together, or temporarily melting the surface of the coil 12 to embed part of the axially oriented element 30.

[0032] An advantage of securing the axially oriented element 30 to both ends 24 and 26 of the coil 12 is that the axially oriented element 30 can function as a stretch-resistant member, which prevents the first end 24 of the coil 12 from being pulled too far from the second end 26. The axially oriented element 30 can also be pre-stretched before it is secured to the ends of the coil 12, to thereby provide some degree of compression within the coil 12. Examples of stretch-

resistant members are described in U.S. Patent Nos. 6,193,728, 6,013,084, 6,004,338, 5,853,418, 5,833,705, 5,582,619.

[0033] In alternative embodiments, instead of securing to both ends of the coil 12, the axially oriented element 30 can be secured to the coil 12 at one of the
5 ends 24 and 26 of the coil 12 or at one or more points along a length of the coil 12 by a suitable adhesive or by wrapping around one or more windings 18 of the coil 12. In another embodiment, the axially oriented element 30 is not secured to the coil 12, but is simply disposed within the lumen 20 of the coil 12, or is coupled to the coil 10 by a surface friction, in which case, the surface of the axially
10 oriented element 30 may be textured to improve the coupling force between the axially oriented element 30 and the coil 12.

[0034] The agent carrier 14 preferably has a cross-sectional dimension such that the overall flexibility of the vaso-occlusive device 10 is not significantly impacted. In one embodiment, the cross-sectional dimension of the agent carrier
15 14 is approximately 0.002 inch less than the internal diameter of the coil 12. However any diameter smaller than the coil internal diameter may also be used. If the agent carrier 14 is also used as a stretch-resistant member, the agent carrier 14 should have a minimum cross-sectional dimension such that the agent carrier 14 can have enough strength to provide some degree of tensile resistance
20 to a stretching of the coil 12.

[0035] The agent carrier 14 includes a bioactive material or agent, such as a thrombogenic or a therapeutic agent, that induces a tissue reaction when placed

within a body. Particularly, the agent carrier 14 is made from a bioactive material or agent that is absorbable or biodegradable. When the vaso-occlusive device 10 is placed in a body, the agent carrier 14 dissolves and releases the agent to its surrounding environment. Alternatively, the agent carrier 14 can be made
5 from a non-biodegradable material, in which case, a coating that comprises a bioactive agent is then deposited on a surface of the agent carrier 14. When the vaso-occlusive device 10 is placed within an aneurysm, a body temperature and/or a reaction with a bodily fluid causes the coating to degrade or dissolve, thereby releasing the bioactive agent.

10 [0036] Notably, the bioactive agent may be incorporated within the agent carrier, e.g., in a cavity, or dispersed within the material comprising the agent carrier itself, such material being either absorbable or non-absorbable.

[0037] Preferably, the bioactive agent is a type which elicits a tissue reaction that leads to rapid in-growth of fibro-cellular tissue, thereby stabilizing the
15 occlusion of the aneurysm without compromising blood flow in the native vasculature. An advantage of placing the agent carrier 14 within the lumen 20 of the coil 12 is that an exterior of the coil 12 is unaffected by the bioactive material during delivery of the coil 12. That is, the bioactive material would not increase a friction between the coil 12 and a delivery tool, and would not cause the coil 12 to
20 be adhered to the delivery tool or to a packaging.

[0038] Examples of materials that can be included in the agent carrier 14 include homopolymers or copolymers comprising in part: polyesters, acrylics,

polyethers, polysiloxanes, polyurethanes, polycarbonates, and other biocompatible polymers. Biodegradable or absorbable materials may also be used in the agent carrier and/or as the bioactive agent and include, but are not limited to, synthetic polymers, polysaccharides, and proteins. Suitable polymers may include, for example, polyglycolic acid, polylactic acid, polycaprolactone, polyhydroxyalkanoates (such as polyhydroxybutyrate and polyhydroxyvalerate), polydioxanone, poly(trimethylene carbonate), polyanhydrides, poly(g-ethyl glutamate), poly(DTH iminocarbonate), poly(bisphenol A iminocarbonate), polyarylates, polyamino acids and copolymers or mixtures thereof.

[0039] In addition, or alternatively, proteins may be used, such as collagen, elastin, caesin, fibrin, fibrinogen, fibronectin, vitronectin, laminin, silk, and/or gelatin. In addition or alternatively, polysaccharides may be used, such as chitin, chitosan, cellulose, alginate, hyaluronic acid, and chondroitin sulfate. Many of these materials are commercially available. Fibrin-containing compositions are commercially available, for example from Baxter Healthcare. Collagen-containing compositions are commercially available, for example, from Cohesion Technologies, Inc., of Palo Alto, California. Fibrinogen-containing compositions are described, for example, in U.S. Patent Nos. 6,168,788 and 5,290,552. As will be readily apparent, absorbable materials may be used alone or in any combination with each other. The absorbable material may be a mono-filament or multi-filament strands or a tube.

[0040] The materials that comprise the carrier can themselves be bioactive.

These materials in their unaltered or in a degraded form may stimulate a biological reaction that ultimately results in the formation of fibro-cellular tissues.

For example, certain polymers such as bioabsorbable polymers or certain

5 polyesters can illicit an inflammatory reaction; certain proteins such as fibrinogen or collagen can illicit a thrombogenic reaction; and other proteins such as silk can illicit an immune response.

[0041] Other examples of bioactive materials that can be included in the agent carrier 14 include cytokines; extracellular matrix molecules (e.g., collagen, fibrin,

10 or decellularized animal tissues); matrix metalloproteinase inhibitors; trace metals

(e.g., copper); other molecules that may stabilize thrombus formation or inhibit

clot lysis (e.g., proteins, including Factor XIII, α_2 -antiplasmin, plasminogen

activator inhibitor-1 (PAI-1), and the like); and their functional fragments (e.g., the

P1 or P2 epitopes of fibrin). Examples of cytokines that may be used alone or in

15 combination with other compounds may include basic fibroblast growth factor

(bFGF), platelet derived growth factor (PDGF), vascular endothelial growth factor

(VEGF), transforming growth factor beta (TGF- β), and the like. Cytokines,

extracellular matrix molecules, matrix metalloproteinase inhibitors, and thrombus

stabilizing molecules are commercially available from several vendors, such as

20 Genzyme (Framingham, MA), Genentech (South San Francisco, CA), Amgen

(Thousand Oaks, CA), R&D Systems, and Immunex (Seattle, WA).

[0042] Additionally, bioactive polypeptides that may be synthesized recombinantly as the sequence of many of these molecules are also available, for example, from the GenBank database. Thus, the agent carrier 14 may include use of DNA or RNA encoded bioactive molecules. Furthermore, 5 molecules having similar biological activity as wild-type or purified cytokines, extracellular matrix molecules, matrix metalloproteinase inhibitors, thrombus-stabilizing proteins (e.g., recombinantly produced or mutants thereof), and nucleic acid encoding these molecules may also be used. The amount and concentration of the bioactive materials that may be included in the composition 10 of the agent carrier 14 may vary depending upon the specific application. It will be understood that any combination of materials, concentration, and/or dosage may be used, so long as it is not harmful to the subject.

[0043] The structural materials that comprise the carrier can themselves be the bioactive agent. These materials in their unaltered or in a degraded form may 15 stimulate a biological reaction that ultimately results in the formation of fibro-cellular tissues. For example, certain polymers such as bioabsorbable polymers or certain polyesters can illicit an inflammatory reaction; certain proteins such as fibrinogen or collagen can illicit a thrombogenic reaction; and other proteins such as silk can illicit an immune response.

20 [0044] In alternative embodiments, instead of being made from a bioactive material, the agent carrier 14 is made from a material that adheres or absorbs a bioactive agent. For examples, the agent carrier 14 may include one or more

polymer filaments, a sponge, a cloth, a hydrogel, or other materials that are capable of absorbing or adhering a bioactive agent. In this case, the agent carrier 14 is used to deliver the bioactive agent, which will diffuse out of the agent carrier 14 into the surroundings when placed in an aneurysm.

5 [0045] The bioactive agent may also be disposed within the carrier, e.g., wherein the carrier has a sealed reservoir containing the agent, or wherein the agent is dispersed within the material comprising the container. In such embodiments, the agent will diffuse out of the carrier. The selected agent preferably elicits a tissue reaction that leads to rapid in-growth of fibro-cellular
10 tissue, thereby stabilizing the occlusion of the aneurysm. The agent may include any of the materials described previously. The agent may also include drugs, proteins, cells, genetic modifiers, inflammatory agents, immuno-agonistic agents (e.g. Freund's adjuvant or squalene), clot stabilizer, clot activators (e.g. thrombin or Factor XIII), cellular materials (e.g. concentrated blood products, fibroblasts,
15 smooth muscle cells, progenitor cells, genetically engineered cells that secrete a particular bioactive protein), viral vectors, or plasmids.

[0046] In-Situ Stiffening of Vaso-Occlusive Device

[0047] In accordance with another aspect of the invention, the vaso-occlusive device 200 of **FIG. 2** is provided with an active element 214 configured
20 to provide in-situ stiffening of the coil 200. The coil 212 is made from a linear element 216, such as a wire, which preferably has a circular cross-sectional shape. Alternatively, the linear element 216 of the coil 212 may have a

rectangular, triangular, other geometric cross-section, or an irregular shaped cross-section.

[0048] The coil 212 includes one or more loops or windings 218 formed by the linear element 216. The loops 218 define a central lumen 220 in which the active element 214 is placed. Any of the materials described previously with reference to the coil 12 is also suitable for construction of the coil 212. In the illustrated embodiment, the vaso-occlusive device 200 has an overall diameter or cross-section which is preferably in the range of 0.010 to 0.023 inches. However, the vaso-occlusive device 200 may have other diameters and/or cross-sections, as well. The vaso-occlusive device 200 may optionally include one or more end caps 222 secured to a first end 224 and/or a second end 226 of the coil 212.

[0049] As shown in **FIG. 2**, the active element 214 includes an axially oriented element 230 having a substantially rectilinear or a curvilinear (less than 360°) configuration along a length of the vaso-occlusive device 200. Again, in the case of more complex coil shapes, the active element could mirror the shape of the coil. The axially oriented element 30 is located within the lumen 220 of the coil 212 and is secured to the ends 224 and 226 or the end caps 222 of the coil 212. The securing may be accomplished by any of the methods described previously.

[0050] In alternative embodiments, instead of securing to both ends 224 and 226 of the coil 212, the axially oriented element 230 can be secured to the coil 212 at one of the ends 224 and 226 of the coil 212 or at one or more points along a length of the coil 212 by a suitable adhesive or by wrapping around one or

more windings 218 of the coil 212. Furthermore, in another embodiment, the axially oriented element 230 is not secured to the coil 212. In this case, the axially oriented element 230 is simply disposed within the lumen 220 of the coil 212, or is coupled to the coil 210 by a surface friction, in which case, the surface
5 of the axially oriented element 230 may be textured to improve the coupling force between the axially oriented element 230 and the coil 212.

[0051] The active element 214 is configured to undergo a reaction that changes a structural characteristic of the vaso-occlusive device 200 when placed in a body cavity. In one embodiment, the active element 214 includes an
10 expansible material, which will expand in size when placed in an aqueous environment within a living mammal. In this case, the cross-section of the active element 214 is configured such that it can expand to a size which is slightly larger than the internal diameter (or the lumen 220) of the coil 212 (**FIG. 3**). In one embodiment, the cross-sectional dimension of the active member 214 is
15 configured such that its swollen cross-sectional dimension is at least equal to 100% - 500%, and more particularly, between 110% - 200%, of the internal diameter of the coil 212. The expanded material imparts a radial stress within the coil 212 to thereby stiffen and stabilize the coil 212 in-situ.

[0052] An example of the expansible material that can be used for
20 construction of the active element 214 is a hydrogel, which is capable of absorbing a desired amount of aqueous fluid. Examples of hydrogels include gels formed from homopolymers, copolymers, and/or network polymers

containing: polyethylene glycol, polypropylene glycol, polyvinyl alcohol, polyvinylpyrrolidone, polyacrylates, polymethacrylates, polyacrylamides, polyethyloxazoline, polysaccharides, mucopolysaccharides, polyaminoacids, carboxy alkyl celluloses, partially oxidized cellulose, hyaluronic acid, dextran, 5 heparin sulfate, chondroitin sulfate, heparin, agar, starch, alginate, fibronectin, gelatin, collagen, fibrin, pectins, albumin, ovalbumin, polyesters of .alpha.-hydroxy acids including polyglycolic acid, poly-DL-lactic, poly-L-lactic acid, polylactones, polyanhydrides, polyorthoesters, polydioxanone, polycaprolactones, poly(delta-valerolactone), poly(gamma-butyrolactone), and 10 combinations thereof. The gel may further comprise a chemical cross-linking agent having two or more reactive groups in order to form chemical bridges between two or more polymeric molecules. Examples of such cross-linking agents include diacrylates, oligoacrylates, dimethacrylates, oligomethacrylates, divinyl ethers, certain cations, and combinations thereof.

15 [0053] The active element 214 can also include a radiopaque material mixed or coated with the hydrogel, or alternatively, include a radiopaque marker secured to the active element 214.

[0054]. Other expansive materials that respond to changes in moisture, ionic strength, temperature, pH, or materials that selectively absorb blood borne 20 substances (e.g. silicones or other polymers which absorb lipids) may also be used. The rate at which the active element 214 expands may be customized, such as by changing a composition of the active element 214, so that placement

and repositioning of the coil 212 may be performed within a period after it has been delivered to a site. The rate at which the active element 214 expands may also be customized or by varying a spacing between the pitch of the coil 212, thereby controlling the amount of bodily fluid that flows into the lumen 220 of the coil 212. The rate may also be controlled by a coating that may be soluble or insoluble which, in either case, limits diffusion of water into the active element.

[0055] In another embodiment, the active element 214 is made from a material that undergoes contraction due to an environmental stimuli, such as moisture, , ionic strength, pH temperature. In this case, the active element 214 is secured to the coil 212 at two points along a length of the coil 212. For example, the active element 214 can be secured to the ends 224 and 226 of the coil 212 using any of the methods described previously. Alternatively, the active element 214 can also be secured to the coil 212 along its length by an adhesive or by wrapping around the loops 218 of the coil 212. When the active element 214 is placed inside a body, it reacts with a stimuli, such as blood or other bodily fluids, and undergoes contraction. Contraction of the active element 214 induces a compressive load on the coil 212 (i.e., the active element 214 compresses the coil 212 between the two points of attachment), which in turn stiffens and stabilizes the coil 212 in-situ.

[0056] Contraction of the active element may be achieved, by way of example, by localized injection of a warm fluid or an ionic solution, provided that the material transition is irreversible. Alternatively, the active element may be

pre-tensioned, wherein prior to use, the device is conditioned with a solvent (e.g., an aqueous ionic solution or polar organic solvent) to cause initial expansion of the element prior to placement inside a body. Once placed in the body, contact with blood causes the solvent to diffuse out of the active element, further causing it to contract.

[0057] Examples of materials that undergoes contraction due to an environmental stimuli includes shape memory alloys and polymers, such as Nitinol™ or polyurethanes and poly(norbornene), which contract (or expand) with changes in temperature. Other, temperature-sensitive contracting materials include protein fibers that undergo thermally induced phase transitions or denaturation at or near body temperature, and thermoresponsive hydrogels, which include polymer gels swollen by aqueous solutions which change volume in response to thermally induced molecular motions that alter the balance of hydrophilic/hydrophobic interactions between the polymer chains and the surrounding aqueous environment. Examples of such hydrogels include ones containing n-isopropyl, acrylamide, chitosan, hyaluronic acid, or poly(ethylene oxide-co-propylene oxide). Examples of materials that contract in response to changes in pH and/or ionic strength include polyelectrolyte hydrogels, which are polymer gels swollen by aqueous solutions which change volume in response to electrostatic interactions between polymer chains and ions in the surrounding aqueous environment. Examples of such hydrogels include ones containing acrylic acid, n-isopropyl acrylamide, amino acids, carboxyl methacrylate, chitosan

or xanthan. Protein fibers that undergo pH induced phase transitions or denaturation at or near body pH may also be suitable for use as a contracting agent, as would polymer gels comprising a biocompatible polymer swollen with a non-aqueous solvent that will diffuse out of the gel upon contact with water (or
5 blood), such as silicones, urethanes, acrylics and polyesters.

[0058] It should be noted that although the vaso-occlusive device 10 has been described as having an agent delivery capability, and the vaso-occlusive device 200 has been described as having an in-situ stiffening capability, the scope of the invention should not be so limited. In an alternative embodiment, a vaso-

10 occlusive device can be configured both to be stiffened in-situ and to deliver a bioactive agent. For example, the vaso-occlusive device 200 configured to be stiffened in-situ can also carry a bioactive agent that causes a tissue reaction when delivered in a body.

[0059] In the previously described embodiments, the agent carrier 14 and the
15 active element 214 have a substantially rectilinear or curvilinear (less than 360°) shape. However, the agent carrier 14 and the active element 214 are not limited to those described previously, and can have other shapes or configurations in alternative embodiments so long as they are located substantially within the central lumen of the coil.

20 [0060] **FIGS. 4-6** each shows a vaso-occlusive device 300 having a coil 302 and an internal element 304 in accordance with alternative embodiments of the present invention. The coil 302 in these figures represents the coil 12 or the coil

212 described previously, and the internal element 304 represents the agent carrier 14 or the active element 214 described previously.

[0061] **FIG. 4** shows a vaso-occlusive device 300(1) that includes a coil 302 and an internal element 304(1) having a shape of a coil. As similarly discussed
5 previously, the internal element 304(1) may be secured to one or both ends of the coil 302, secured to the coil 302 at one or more points along a length of the coil 302, or coupled to the coil 302 by a surface friction.

[0062] **FIG. 5** shows another vaso-occlusive device 300(2) that includes a coil 302 and one or more internal elements 304(2). In this case, the internal element
10 304(2) does not extend approximately from one end to the other end of the coil 302. Instead, the internal element 304(2) only extends along a portion of the length of the coil 302. In the illustrated embodiment, the internal element 304(2) has a shape of a cylinder. However, the internal element 304(2) can also have other shapes or configurations. For examples, the internal element 304(2) can
15 be in a form of a sphere, a cone, a plate, a mesh, or other customized shapes.

FIG. 6 shows a vaso-occlusive device 300(3) that includes an internal element 304(3) having a random shape.

[0063] The vaso-occlusive devices shown in the above-described
embodiments generally have a substantially rectilinear (straight) or a curvilinear
20 (slightly curved, i.e. having less than 360° spiral) relaxed configurations. Such devices may assume folded or bent configurations when they are subjected to an external force (e.g., compressive forces generated when they are pushed against

an object, such as the wall of an aneurysm). The devices may also assume a variety of secondary shapes or relaxed configurations. The space-filling capacity of these vaso-occlusive devices is inherent within the secondary relaxed shape of these devices.

5 [0064] **FIGS. 7-13** illustrate various vaso-occlusive devices 400 that include a coil 402 having a primary shape and a secondary shape. These shapes are simply indicative of the various secondary shapes that may be used, and other shapes may be used as well. While not always shown, the devices 400 illustrated in **FIGS. 7-13** are each provided with an internal element 304, which
10 can be the agent carrier 14 or the active element 214, as discussed previously.

[0065] **FIG. 7** depicts a device 400(1) having a secondary shape of a clover leaf. **FIG. 8** depicts a device 400(2) having a secondary shape of a twisted figure-8. **FIG. 9** depicts a device 400(3) having a flower-shaped secondary shape. **FIG. 10** depicts a device 400(4) having a substantially spherical
15 secondary shape. **FIG. 11** illustrates a device 400(5) having a random secondary shape. **FIG. 12** illustrates a device 400(6) having a secondary shape of a vortex. **FIG. 13** illustrates a device 400(7) having a secondary shape of an ovoid. It should be noted that vaso-occlusive device 400 may also have other secondary shapes, and that it should not be limited to the examples illustrated
20 previously. For example, the vaso-occlusive device 400 may be selectively sized to fill a particular aneurysm or body cavity.

[0066] To make a secondary shaped vaso-occlusive device 400, a coil (i.e., the coil 402) having a primary shape that is substantially rectilinear or curvilinear may be wrapped around a mandrel or other shaping element to form a secondary shape. The coil 402 may be heat treated to shape the coil 402 into the

5 secondary shape. Stable coil designs, and methods of making them, are described in U.S. Patent No. 6,322,576B1 to Wallace et al. It should be noted that forming vaso-occlusive devices into secondary shapes is well known in the art.

[0067] A method of using the previously described vaso-occlusive devices
10 (i.e., device 300) will now be discussed with reference to **FIGS. 14** and **15**. First, a delivery catheter 502 is inserted into the body of a patient. Typically, this would be through a femoral artery in the groin. Other entry sites sometimes chosen are found in the neck, for example, and are in general well known by physicians who practice these types of medical procedures. The delivery catheter 502, which
15 may be a microcatheter or a sheath, may be positioned so that the distal tip 508 of the delivery catheter 502 is appropriately situated, e.g., within the mouth of the body cavity 501 to be treated. The insertion of the delivery catheter 502 may be facilitated by the use of a guidewire and/or a guiding catheter, as is known in the art. In addition, the movement of the catheter 502 may be monitored, for
20 example, using fluoroscopy, ultrasound, and the like.

[0068] Once the delivery catheter 502 is in place, the vaso-occlusive device 300 is then inserted from the proximal end (not shown) of the delivery device

502, and into the lumen of the delivery device 502. This step is not necessary if the vaso-occlusive device 300 is already pre-loaded into the delivery catheter 502. Since the vaso-occlusive device 300 has no secondary shape, the vaso-occlusive device 300 would naturally assume a substantially rectilinear or a curvilinear configuration when disposed within the lumen of the delivery device 502, without being subjected to a substantial stress. For vaso-occlusive devices having secondary shapes, such as the vaso-occlusive devices 400 shown in **FIGS. 7-13**, they may be "bent" to a substantially linear shape while residing within the lumen of the delivery catheter 502, as illustrated in **FIG. 15**.

10 [0069] Referring back to **FIG. 14**, the vaso-occlusive device 300 is preferably advanced distally towards the distal end 508 of the delivery catheter 502 using a core wire or pusher member 504. A plunger 506 may be attached to the distal end of the wire 504 to advance the vaso-occlusive device 300. Alternatively, fluid pressure may also be used to advance the vaso-occlusive device 300 along the delivery catheter 502. The inner diameter of the delivery catheter 502 should be made large enough to advance the vaso-occlusive device 300. On the other hand, the inner diameter of the delivery catheter 502 should not be significantly larger than the overall cross-sectional dimension of the vaso-occlusive device 300 in order to avoid buckling and/or kinking the vaso-occlusive device 300 within the lumen of the delivery catheter 502.

20 [0070] Additional vaso-occlusive devices 300 may also be placed within the body cavity 501 by repeating the relevant steps discussed above. When a

desired number of vaso-occlusive devices has been placed within the body cavity 501, the delivery catheter 502 may be withdrawn from the body cavity 501 and the patient's body. Once the vaso-occlusive devices are deployed in the body cavity 501, an embolic mass is formed therein to occlude the body cavity 501.

5 [0071] Other devices and methods for discharging shaped coils and linear coils into a body cavity may also be used. **FIGS. 16** and **17** each shows a detachable device that can release a vaso-occlusive coil at a specifically chosen time and site.

[0072] **FIG. 16** depicts an embodiment, generally designated 600, having a
10 vaso-occlusive device 602 that may be deployed from a sheath or a catheter 610 through operation of a connective joint 604. The vaso-occlusive device 602 may be any of the devices depicted in **FIGS. 1-13**, i.e., including the agent carrier 14 or the active element 214 (not shown for clarity). Joint 604 has a clasp section 606 that may remain attached to a core wire 612 when the sheath or catheter
15 body 610 is retracted proximally. Joint 604 also may include a second clasp section 608, carried on the proximal end of the vaso-occlusive device 602 and interlocking with clasp section 606 when the assembly is within the sheath 610. When the sheath 610 is withdrawn from about the assembly, the clasp sections may disengage, thereby detaching the vaso-occlusive device 602.

20 [0073] The vaso-occlusive devices described herein may also be detachable by an electrolytic joint or connection, such as described in U.S. Patent Nos. 5,234,437, 5,250,071, 5,261,916, 5,304,195, 5,312,415, and 5,350,397.

[0074] FIG. 17 shows an embodiment, generally designated 660, having a vaso-occlusive device 662 that may be detached using a connective joint 664 that is susceptible to electrolysis. The vaso-occlusive device 662 may be any one of the devices depicted in FIGS. 1-13, and includes the agent carrier 14 or the active element 214 (not shown for clarity). Joint 664 may be made of a metal which, upon application of a suitable voltage to a core wire 668, may erode in the bloodstream, thereby releasing the vaso-occlusive device 662. The vaso-occlusive device 662 may be made of a metal that is more "noble" in the electromotive series than the joint 664. A return electrode (not shown) may be supplied to complete the circuit.

[0075] The region of core wire 668 proximal to the joint is insulated to focus the erosion at the joint. A bushing 666 may be used to connect the distal end of core wire 404 to the proximal end of the vaso-occlusive device 662. To deploy the vaso-occlusive device 662, the vaso-occlusive device 662 attached to the core wire 668 is first placed within a body cavity. An electric current is then applied to the core wire 668 to dissolve the connective joint 664, thereby detaching the vaso-occlusive device 662 from the core wire 668. It should be noted that methods of delivering vaso-occlusive devices by electrolytic disintegration of a core wire joint are well known in the art, and need not be described in further detail. The above described joint and similar joints are described in detail in U.S. Patent No. 5,423,829; 6,165,178, and 5,984,929.

[0076] Although preferred embodiments of the invention are shown and described herein, it would be apparent to those skilled in the art that many changes and modifications may be made thereto without the departing from the scope of the invention, which is defined by the appended claims.

5 [0077] By way of one example, it will be readily apparent that the two main aspects of the invention disclosed and described herein – that of an occlusive device carrying an active element for providing in-situ stiffening of the device after its placement in a selected site in the vasculature, and that of an occlusive device including an agent carrier comprising (or otherwise carrying) a biological
10 agent that elicits a biological reaction inside a body, may be combined in a single embodiment of the invention.